

PROTEIN/PEPTIDE SEQUENCE ANALYSIS SUBMISSION FORM

BILLING INFORMATION

| RESEARCHER INFORMATION | |
|------------------------|--|
| | |

| Date: | P.O./Credit Card #: |
|-------------------------|---------------------------------|
| Principal Investigator: | Exp. Date (if applicable): CVV: |
| Department/Company: | Department/Company: |
| Phone: Fax: | Phone: Fax: |
| Email: | Email: |
| SHIPPING ADDRESS: | BILLING ADDRESS: |
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Sample Information:

Please provide as much information as possible:

| 1. | SAMPLE AMOUNT: moles; or | r micrograms | |
|----|---|---|----------|
| | A. Recommend minimum amount of samples is 100 pMo | ples. | |
| | Example: 100 pmoles of a protein (M.W. 10,000) would | weigh 1.0 μg | |
| | B. If less than 100 pmoles of protein sample, there are NO | GUARANTEES of adequate sequencing results. | |
| 2. | Number of cycles per sample: | | |
| 3. | Is sample in SOLUTION or DRY: | | |
| | A. If sample is in solution, what is the sample stored in? | | |
| 4. | METHOD(s) USED TO OBTAIN THE SAMPLE: | | |
| | A. For samples electroblotted to PVDF: What membrane | was used? | |
| | Immobilon-P (.45 micron) (Millipore) | Problot (.1 micron) (ABI) | |
| | Westran (.45 micron) (Schleicher & Schuell) | Trans-Blot (.1 micron) (Biorad) | |
| | Immobilon-PSQ (.1 micron) (Millipore) | Fluoratrans (.1 micron) Pall Corp) | |
| | B. It is possible to sequence directly from a membrane: I | PVDF & Immobilon | |
| 5. | ESTIMATED PURITY OF THE SAMPLE: | | |
| | A. Method(s) used to obtain Purity: | | _ |
| | B. If your sample was collected on a HPLC, please at and wavelength | ttach the chromatogram with gradient, solvents, col | lumn |

| 6. | N-TERMINUS BLOCKED? Yes Do not know No |
|-----|--|
| | A. If the peptide/protein is blocked, it will NOT be able to be sequenced, unless cleaved by enzyme(s) first. |
| | B. The only way to determine if the peptide/protein is blocked is to: 1. Conduct a tryptic digest apriori 2. Attempt to sequence, and see what happens |
| 7. | DOES SAMPLE CONTAIN? Tris Salts Amino Containing Buffers |
| | Tris-Glycine Detergents |
| | 1. If yes, name of detergent: |
| | 2. % of detergent in the sample: |
| | A. Sequencing is hindered by the presence of any salt. Amino containing buffers, and some detergents. |
| | B. De-salting is best accomplished by double dialysis treatment |
| 8. | ARE THERE ANY KNOWN MODIFIED AMINO ACIDS? |
| | Phosphorylations: Conjugations: Cysteine Modified: If yes, what derivative?: |
| 9. | ENZYME TREATMENT: Yes No What enzyme? Cleavage sites |
| | Phosphorylations: Conjugations: Cysteine Modified: If yes, what derivative?: |
| 10. | PROTEIN SEQUENCE KNOWN: Yes 🗌 No 📄 (Please attach) |
| 11. | AMINO ACID ANALYSIS PERFORMED? Yes 🗌 No 📄 (Please attach) |

Note: Proper sample preparation is crucial for optimal results. Important parameters include concentration and volume of sample, as well as the presence and concentration of detergents, glycerol, buffers and other salts. Requirements vary depending on the analysis requested. Do not make any assumptions about sample preparation requirements.Customer samples will be discarded unless otherwise specified. Consult with our technical support at support@biosyn.com or call 800-227-0627 for detailed

Label sample to be sequence properly and return to:

Bio-Synthesis Inc. Analytical Service Coordinator 612 E. Main Street Lewisville, TX 75057

