

Using an RNA aptamer probe for flow cytometry detection of CD30-expressing lymphoma cells

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Aptamers are small molecular ligands composed of short oligonucleotides that bind targets with high affinity. In contrast to antibodies, as synthetic oligonucleotides, aptamers have lower production costs and elicit no antigenic reactions. Therefore, aptamers are potential agents for disease diagnosis and treatment. In this study, we validate a fluorescently labeled RNA aptamer, which has been reported to bind specifically to mouse CD30 proteins in solution, for human CD30 protein recognition on intact cells. The aptamer probe was tested with cultured anaplastic large cell lymphoma and Hodgkin's lymphoma cells that express high levels of CD30. Flow cytometry and fluorescence microscopy showed specific and sensitive binding of the aptamer probe to CD30-expressing lymphoma cells at low concentrations (0.3 nM). Studies performed on multiple cell lines and nuclear cells from healthy donors confirmed that the CD30 aptamer and anti-CD30 antibody, the standard clinical probe, recognized the same set of cells. The potential application of multicolor flow cytometry analysis using the CD30 aptamer probe and antibodies was also shown. In conclusion, the developed CD30 aptamer probe could act as a replacement and/or a supplement for antibodies in the diagnosis of the CD30-expressing lymphomas.

Laboratory Investigation (2009) 89, 1423–1432; doi:10.1038/labinvest.2009.113; published online 12 October 2009

KEYWORDS: aptamer probe; CD30 expression; cell binding; flow cytometry analysis; lymphoma diagnosis; RNA oligonucleotide

In the past two decades, a new class of small molecule ligands composed of short oligonucleotides (usually 20–50 bases), known as 'aptamers', has been widely investigated.^{1–7} Use of the systematic evolution of ligands by EXponential amplification technique has led to a variety of RNA or single-stranded DNA aptamers that have high binding affinity to their target molecules at pico- to nanomolar levels.^{7,8} Aptamers can virtually bind to all types of targets, including ions, toxins, drugs, low-molecular-weight ligands, peptides, proteins, and cells. A recent study that used the extracellular domain of the receptor activator of nuclear factor- κ B (RANK) as a target identified an RNA aptamer that specifically binds to the tumor necrosis factor (TNF) receptor family proteins.⁹ Interestingly, among its binding targets, this aptamer binds to mouse CD30 proteins with the highest affinity, more than 1000 times higher than other tested proteins among the TNF receptor family members in solution.

CD30 antigen is a transmembrane receptor that is normally expressed by activated T cells.¹⁰ Abnormal expression of CD30 has been identified in tumor cells of anaplastic large cell lymphoma (ALCL) and Hodgkin's lymphoma.^{11,12} Detection of CD30 expression on tumor cells is a requirement for diagnosis of these lymphomas and for differentiating them from other types of tumors.¹³ To date, the anti-CD30 antibody is the only probe that has been validated to detect cellular CD30 expression in clinical settings.¹⁴ It is known that the binding of cell surface CD30 to its ligands could trigger complicated cellular responses and might be involved in the pathogenesis of the CD30-expressing neoplasms.^{15–17} Thus, cellular CD30 receptor targeting represents a new therapeutic strategy for both diagnosing and treating ALCL.^{18–22} In contrast to protein antibodies, aptamers are small, synthetic oligonucleotides, and therefore they are not antigenic *in vivo* and show better tissue/cell penetration.^{7,23} To validate the potential usage of aptamers in lymphoma

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Received 29 May 2009; revised 10 August 2009; accepted 25 August 2009

diagnosis, we tested the previously identified RNA aptamer as a specific probe to detect the human CD30-expressing lymphoma cells. Furthermore, the usage of multicolor immunophenotyping was shown by combining the fluorescently labeled aptamer probe and antibodies.

MATERIALS AND METHODS

Cell Culture, Cell Preparation, and Reagents

In this study, the human systemic ALCL cell lines Karpas 299 and SUDHL-1 were used in collaboration with Dr Mark Raffeld at the National Cancer Institute/National Institutes of Health. The human acute T-cell leukemia cell line Jurkat, human myeloid leukemia cell line K562, and human myeloma cell line RPMI8226 were purchased from the American Type Culture Collection (ATCC, Manassas, VA, USA). Human Hodgkin's lymphoma cell lines, L540, HDLM2, L428, and KMH2 were used in collaboration with Dr Barbara Savoldo at the Baylor College of Medicine (Houston, TX, USA). Cells were routinely cultured in RPMI-1640 medium (Mediatech, Manassas, VA, USA) supplemented with 10% fetal bovine serum (FBS; Invitrogen, Carlsbad, CA, USA), 100 units/ml penicillin, and 100 µg/ml streptomycin (Atlanta Biologicals, Lawrenceville, GA, USA) at 37°C in a 5% CO₂ atmosphere.

For preclinical evaluation, nuclear cells were prepared from bone marrow aspirates from healthy donors under an Institutional Review Board approved protocol ($n=10$). In brief, 1 ml anticoagulant-treated bone marrow specimens were centrifuged at 500 g for 10 min, and nuclear cells within the Buffy coat between the serum and cell pellets were collected. Contaminated red blood cells (RBCs) were lysed by 6 ml of RBC lysis buffer (Cell Signaling Technology, Inc., Danvers, MA, USA) at room temperature for 10 min. Unlysed nuclear cells were then washed with 2 ml phosphate-buffered saline (PBS, pH 7.4) and resuspended in cold PBS. In addition, to prepare fresh lymphoma cells, the patients' lymph nodes involved by follicular lymphoma ($n=5$) were digested by vigorous cutting and repeated pipetting to generate a cell suspension in cold PBS. The cells were then filtered through a 100 µm cup filcon (BD Mediatech, San Jose, CA, USA) to obtain a single-cell suspension.

Preparation of RNA Aptamer Probe Specific for CD30

According to the reported aptamer sequences,⁹ a 39-mer nucleotide RNA aptamer was synthesized with one minor modification: 5'-gauUCGUAUGGGUGGGAUCGGGAAGGG CUACGAACaccg-3' (Bio-Synthesis, Inc., Lewisville, TX, USA). The synthesized aptamer probe contains the most essential functional motif and has the highest affinity to mouse CD30 molecules in solution.⁹ To enhance its resistance to nucleases, the first three nucleotides on each end of the aptamer were synthesized with 2'-O-methyl modification (indicated by lower case letters). The predicted 3D structure of the aptamer is shown in Figure 1a. For tracking purposes, the synthesized aptamers were labeled with the fluorochrome cyanine dye, Cy5, at the 5' end. The

aptamers were purified by high performance liquid chromatography and stored at -20°C until use. To rule out nonspecific cell binding, a control 39-mer RNA probe was synthesized with a random sequence and also labeled with the Cy5 reporter: 5'-gau- random rN ($n=33$)-ccg-3' (Figure 1a).

Cell Binding Assay and Flow Cytometry Analysis

To determine the optimal cell binding conditions for the synthesized CD30 aptamer probes, Karpas 299 cells that express CD30 and Jurkat cells that do not express CD30 were used. Cells (5×10^5 /assay) were incubated with the CD30 aptamer probes (0.3 nM) in 100 µl of cell binding buffer (see below) at room temperature for 20 min and then washed once with binding buffer without bovine serum albumin (BSA). The cells were analyzed by an LSRII flow cytometer (BD Biosciences, San Jose, CA, USA) with FlowJo (v7.0) software. To validate the optimal cell binding conditions, different concentrations of CaCl₂, MgCl₂, and BSA (Sigma-Aldrich, St. Louis, MO, USA) in PBS were tested as indicated in Figure 2. After confirmation, the optimized binding buffer (PBS, pH 7.4) containing 1 mM MgCl₂ and 0.1% BSA was used throughout the remaining experiments. To determine the optimal concentration of the CD30 aptamer probe, a dosing experiment was performed using different concentrations of aptamer probes (0–3 nM) as described in Figure 2. A standard concentration for cell binding assays of the 0.3 nM CD30 aptamer was selected for all additional experiments.

To validate the binding specificity of the CD30 aptamer, various cultured lymphoma and leukemia cell lines were used as listed in Table 1. Cells were incubated with the Cy5-labeled CD30 aptamer, as described above, or the fluorescein isothiocyanate (FITC)-labeled anti-CD30 antibody (BD Biosciences), a standard probe for examining CD30-expressing cells in clinical laboratories. For cell staining, the 0.3 nM CD30 aptamer or 1:20 diluted anti-CD30 antibody was added to 5×10^5 cells in 100 µl binding buffer and incubated at room temperature for 20 min. After washing with binding buffer without BSA, probe or antibody binding was detected by flow cytometry analysis. For further validation, nuclear cells from healthy donor bone marrow ($n=10$), which included granulocytes, monocytes, lymphocytes, and nuclear RBCs, and fresh lymphoma cells from patients' lymph nodes involved by follicular lymphoma ($n=5$) were stained with the CD30 aptamer or anti-CD30 antibody.

To examine whether the CD30 aptamer and CD30 antibody compete for the same binding site on the CD30 protein, cultured Karpas 299 cells (5×10^5) were co-stained with the Cy5-labeled CD30 aptamer (0.3 nM) and FITC-labeled CD30 antibody (1:20 dilution) for 20 min. Cell staining by each probe was evaluated by flow cytometry analysis.

To determine whether the CD30 aptamer probes could be combined with other antibodies to produce multicolor cell staining, phycoerythrin (PE)-labeled anti-CD25 antibody, FITC-labeled anti-CD30 antibody, and AmCyan-labeled anti-

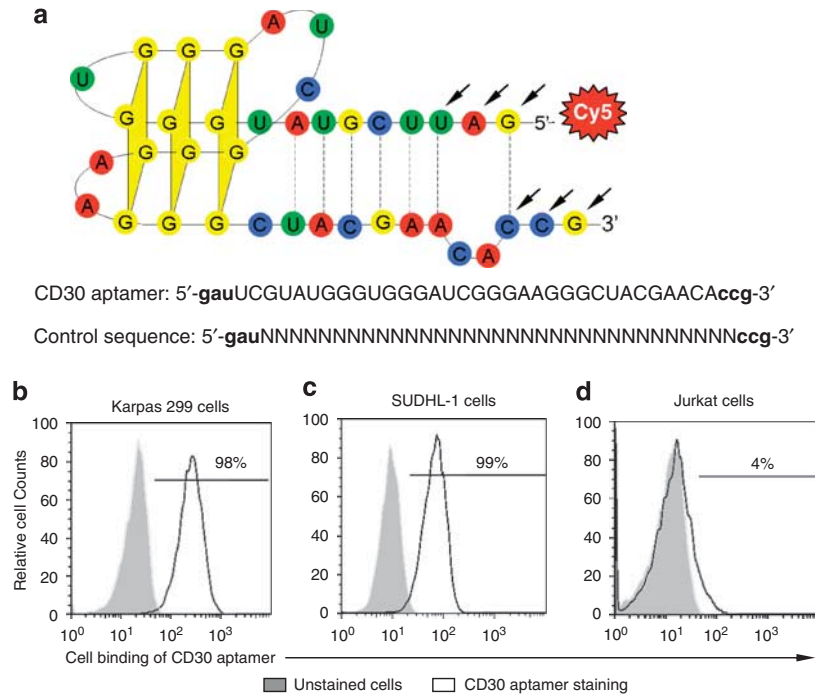


Figure 1 Specific binding of the synthesized CD30 aptamer to human lymphoma cells. **(a)** The synthesized CD30 aptamer sequence and its predicted structure. A 39-mer RNA aptamer was synthesized according to the published sequence (9) and labeled with fluorochrome Cy5 at its 5'-end. To improve its nuclease resistance, the first three nucleotides on each end were synthesized with 2'-O-methyl modification (arrows). To rule out nonspecific cell binding, a control 39-mer RNA probe was synthesized with a random sequence and also labeled with the Cy5 reporter as illustrated. **(b-d)** Cell binding was assessed by flow cytometry analysis. Cultured human ALCL cells (Karpas 299 and SUDHL-1, which express a high level of CD30 proteins) and Jurkat cells that do not express CD30 were incubated with the 0.3 nM CD30 aptamer, and the resultant aptamer–cell binding was detected by flow cytometry analysis. Gray peaks represent unstained cells, and open peaks represent cells that were stained with the CD30 aptamer.

CD45 antibody from BD Biosciences were tested. Cultured Karpas 299 cells, SUDHL-1 cells, and Jurkat cells (5×10^5) were stained with mixed probes, including the 0.3 nM Cy5-labeled aptamer, 1:20 anti-CD25 antibody, 1:20 anti-CD30 antibody, and 1:20 anti-CD45 antibody at room temperature for 20 min. Cells were washed once, and binding of the aptamer and antibodies to cells was detected by an LSRII flow cytometer with four-color channels.

To detect ALCL cells in a heterogeneous cell specimen, fresh nucleated cells (5×10^5) from the bone marrows of healthy donors were prepared by lysing the RBCs using RBC lysis buffer as described above. A small amount of cultured Karpas 299 cells (~ 0.5 –13% of total cellularity) were added into the marrow cell specimens and the mixed cells were incubated simultaneously with the Cy5-labeled CD30 aptamer, AmCyan-labeled anti-CD45 antibody, and FITC-labeled anti-CD30 antibody for 20 min as described above. Stained cells were washed once and flow cytometry analysis was performed using an LSRII flow cytometer with three-color channels. The individual cellular populations in the cell mixture were separated and gated according to their distribution on the side scatter (SSC) vs CD45 panels following standard protocol for clinical flow cytometry (Figure 7a). Cellular CD30 expression in each gated cell population was examined using both the CD30 aptamer probe and anti-

CD30 antibody, and the numbers of detected CD30-positive Karpas 299 cells were compared.

Cell Staining Analysis of CD30 Aptamer Probes by Fluorescence Microscopy

Specific CD30 binding of aptamer was further examined using fluorescence microscopy. In this experiment, CD30-negative Jurkat cells (1×10^6) were pre-labeled with 25 nM of carboxyfluorescein diacetate succinimidyl ester (CFSE, Invitrogen, Eugene, OR, USA) in 1 ml of PBS in the dark at 37°C for 15 min.²⁴ The cells were then washed with PBS once and recultured in 4 ml fresh RPMI 1640 medium containing 10% FBS for 30 min before use. A cell mixture was prepared by mixing the CFSE-labeled Jurkat cells (5×10^5) with an equal number of fresh Karpas 299 cells in binding buffer. Cy5-labeled CD30 aptamer probe (0.3 nM) was added to the cell mixture and incubated at room temperature for 20 min. After washing, slides of cell smears were prepared and examined under a fluorescent microscope.

RESULTS

RNA Aptamer Specifically Binds to Cultured Human CD30-Expressing Lymphoma Cells

To test whether the reported RNA aptamer recognizes proteins of the mouse TNF receptor family and is able to bind to

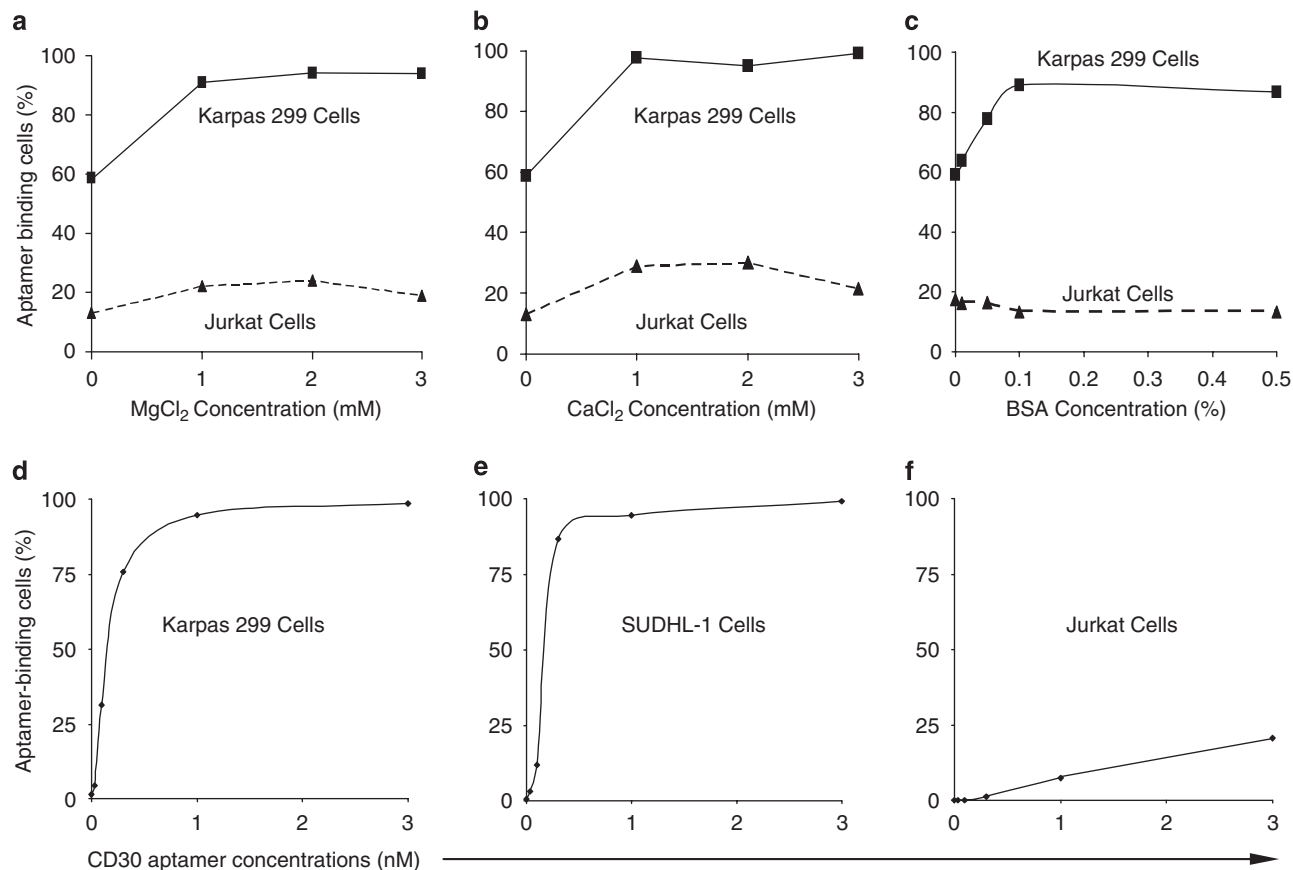


Figure 2 Optimal cell binding conditions for the CD30 aptamer. (a) Effect of MgCl₂ on cell binding. Karpas 299 cells were incubated with the CD30 aptamer in PBS (pH 7.4) with varying concentrations of MgCl₂ ranging from 0 to 3 mM. Resultant aptamer–cell binding was detected by flow cytometry analysis. (b) Effect of CaCl₂. The cell binding assay was repeated with varying concentrations of CaCl₂ ranging from 0 to 3 mM. (c) Effect of BSA. The cell binding assay was repeated with varying concentrations of BSA ranging from 0 to 0.5% (wt/wt). (d–f) Dosage study of the CD30 aptamer. Cultured Karpas 299 (d), SUDHL-1 (e), and Jurkat (f) cells were incubated with increasing concentrations of CD30 aptamer from 0 to 3 nM as indicated. The resultant aptamer–cell binding (%) was detected by flow cytometry analysis.

human CD30 on intact cells, a 39-mer RNA aptamer was synthesized according to the published sequence⁹ and labeled with fluorochrome Cy5 (Figure 1a). The first three nucleotides on each end of the aptamer were replaced with 2'-O-methyl nucleotides for better nuclease resistance (indicated by arrows). To assess cell binding, Karpas 299 and SUDHL-1 cells, which are human ALCL cell lines that characteristically express high levels of CD30 on their surfaces, were tested. Jurkat cell, a human T-cell leukemia cell line that does not express CD30, was used as a negative control. Cells were incubated with the CD30 aptamer probes (0.3 nM) in binding buffer for 20 min at room temperature, and resultant cell binding of the CD30 aptamer probe was quantitated by flow cytometry analysis. As shown in Figures 1b–d, the CD30 aptamer probes only bound to Karpas 299 and SUDHL-1 cells, but not to Jurkat cells. More than 98% of the Karpas 299 and SUDHL-1 cells were labeled with the probes.

To optimize binding conditions for the CD30 aptamer, PBS buffer (pH 7.4) with varying concentrations of MgCl₂, CaCl₂, and BSA were tested. The presence of Mg or Ca ions enhanced CD30 aptamer binding to cultured Karpas 299 cells

with a maximal effect at a final concentration of 1 mM Mg²⁺ or Ca²⁺ (Figures 2a and b), but had little effect on nonspecific aptamer binding to Jurkat cells. The addition of BSA resulted in a significant increase in CD30 aptamer binding to Karpas 299 cells with a maximal effect at 0.1% concentration, and also reduced nonspecific binding to Jurkat cells (Figure 2c), thereby increasing the overall specificity of CD30 aptamer–cell binding. Similar CD30 aptamer–cell binding patterns were observed with SUDHL-1 cells (data not shown). Thus, PBS buffer containing 1 mM MgCl₂ and 0.1% BSA was used as the standard binding buffer in all further experiments. Thereafter, a dosing study was performed to determine optimal concentration of CD30 aptamer for cell binding. As shown in Figures 2d and 2e, CD30 aptamer had a high affinity to Karpas 299 and SUDHL-1 cells and reached saturation after 1 nM. In contrast, the CD30 aptamer showed minimal nonspecific binding to Jurkat cells (Figure 2f).

To rule out nonspecific binding of RNA aptamer to cultured cells, a 39-mer randomized control RNA aptamer probe was synthesized (Figure 1a) and tested. As expected, flow cytometry analysis showed that this randomized control probe had little

to no binding capacity to Jurkat and Karpas 299 cells (Figures 3a–d). To further validate cell-specific binding of the CD30 aptamer, fluorescent microscopic examination was carried out.

Table 1 Comparison of anti-CD30 antibody and CD30 aptamer binding to various cell types

Tested cells	Anti-CD30 antibody	CD30 aptamer
<i>Cell lines</i>		
Jurkat (T-cell lymphoma)	–	–
K562 (myeloid leukemia)	–	–
RPMI8226 (multiple myeloma)	–	–
SUDHL-1 (ALCL)	+	+
Karpas 299 (ALCL)	+	+
L-540 (Hodgkin lymphoma)	+	+
HDLM2 (Hodgkin lymphoma)	+	+
L-428 (Hodgkin lymphoma)	+	+
KMH2 (Hodgkin lymphoma)	+	+
<i>Cells from lymphoma tissues*</i>		
Follicular lymphoma cells	–	–
<i>Cells from bone marrows**</i>		
Granulocytes	–	–
Monocytes	–	–
Lymphocytes	–	–
Nuclear RBCs	–	–

*n = 5; **n = 10. RBCs, red blood cells.

For this purpose, Karpas 299 cells were mixed with CD30-negative Jurkat cells that had been pre-stained with CFSE. The cell mixtures were then incubated with the Cy5-labeled CD30 aptamer probe. Fluorescent microscopy (Figure 3e) showed that the CD30 aptamer probe specifically bound to Karpas 299 cells (the cells with unstained cytoplasm and red fluorescent signal on surface), but did not bind to the CFSE-labeled Jurkat cells (the cells with green fluorescent cytoplasm and no red fluorescent signal on the cell surface).

Identical Cell Binding Specificity and Sensitivity by the CD30 Aptamer Probe and Anti-CD30 Antibody

Currently, antibody staining is the standard histological method to detect CD30 expression in clinical tissues. To confirm specific cell binding, the aptamer was checked with a series of CD30-expressing cell lines, including human ALCL cells (SUDHL-1 and Karpas 299) and Hodgkin’s lymphoma cells (L540, HDLM2, L428, and KMH2), and CD30-negative cell lines, including T-cell leukemia cells (Jurkat), myeloid leukemia cells (K562), and multiple myeloma cells (RPMI8226). Cells were incubated with the 0.3 nM Cy5-labeled CD30 aptamer or FITC-labeled anti-CD30 antibody in binding buffer for 20 min. Cell binding by the aptamer and the antibody was analyzed by flow cytometry. Both the CD30 aptamer probe and anti-CD30 antibody showed identical cell binding patterns. Strong staining was seen with all tested CD30-expressing cells (SUDHL-1, Karpas 299, L340, HDLM2, L428, and KMH2 cells), whereas little or no staining was seen with CD30-negative cells (Jurkat, K562, and RPMI8226 cells) (Figure 4; Table 1).

For further validation, nuclear cells, including granulocytes, monocytes, lymphocytes, and nuclear RBCs, from marrow aspirates of healthy donors (n = 10) were prepared. Cells from fresh lymph node tissue sections of follicular lymphoma patients (n = 5) were also prepared as described

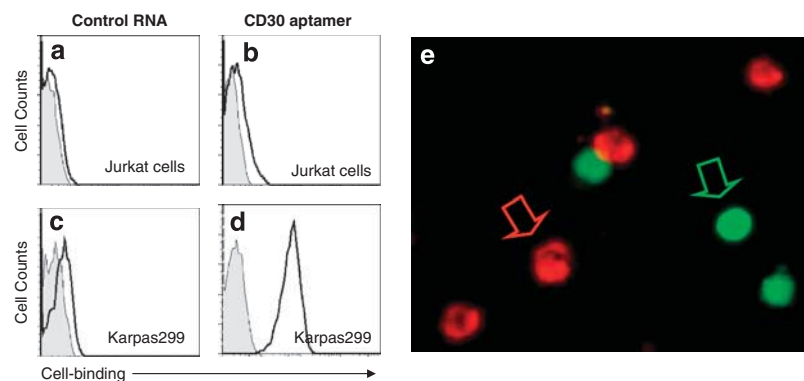


Figure 3 Specific cell staining by the CD30 aptamer. (a–d) Cell binding assays. To rule out nonspecific cell binding, a 39-mer control RNA probe with a random sequence was synthesized (Figure 1a) and incubated with cultured lymphoma cells. The resultant aptamer–cell binding was examined by flow cytometry. Gray peaks represent unstained cells, and open peaks represent cells that were stained with the aptamer probes as indicated. (e) Fluorescent cell staining by the CD30 aptamer. Jurkat cells were pre-labeled with the fluorescent dye CFSE and mixed with fresh, untreated Karpas 299 cells. Cell mixtures were then incubated with the Cy5-labeled CD30 aptamer, and the resultant aptamer–cell staining was examined under a fluorescent microscope. Pre-labeled Jurkat cells were identified by green fluorescence and are indicated by the green arrow, and Karpas 299 cells that were specifically stained by CD30 aptamer were identified by red fluorescence and are indicated by the red arrow.

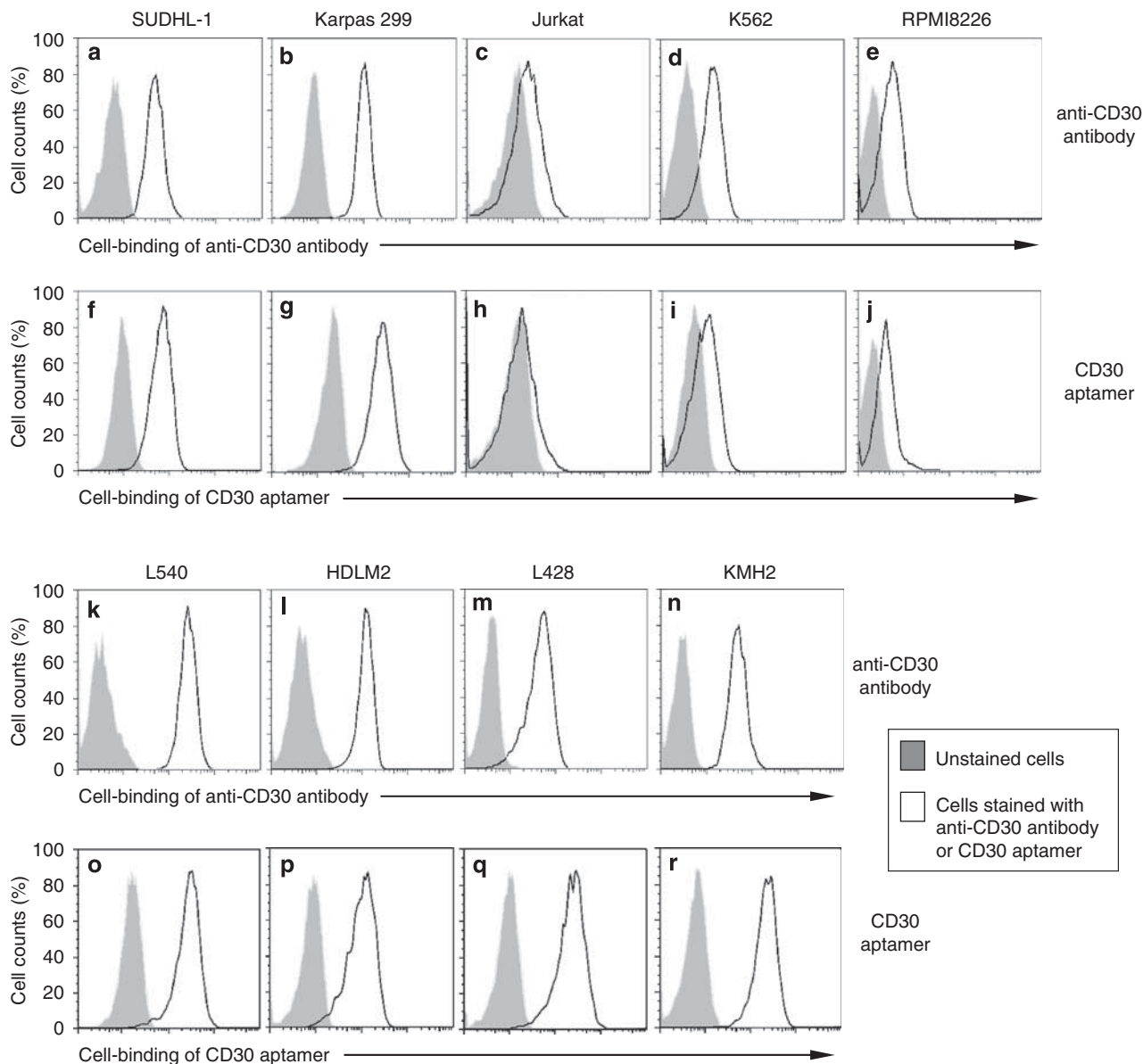


Figure 4 Identical cell binding patterns of the CD30 aptamer and anti-CD30 antibody. Cultured human lymphoma and leukemia cells (as indicated) were incubated in binding buffer containing the FITC-labeled anti-CD30 antibody (a–e and k–n) or Cy5-labeled CD30 aptamer (f–j and o–r) for 20 min. Cell binding was examined by flow cytometry analysis. Gray peaks represent unstained cells, and open peaks represent cells that were stained by the aptamer or antibody.

in 'Materials and Methods.' The fresh cells were probed by the AmCyan-labeled anti-CD45 (common leukocyte antigen) antibody and Cy5-labeled CD30 aptamer. In the control group, the FITC-labeled anti-CD30 antibody was used instead of the CD30 aptamer. Stained cells were analyzed by flow cytometry, and populations of granulocytes, monocytes, and lymphocytes were gated according to their distribution on the side scatter vs CD45 panels. Subsequently, cell binding of the CD30 aptamer to the gated individual cell populations was evaluated and compared with that of the anti-CD30 antibody. The CD30 targeting aptamer and the anti-CD30 antibody showed identical cell staining patterns (Table 1).

Competition Between the CD30-Specific Aptamer and Anti-CD30 Antibody

To examine whether the CD30 aptamer probe and anti-CD30 antibody competitively bind to the CD30 protein, cultured Karpas 299 cells were simultaneously incubated with the Cy5-labeled CD30 aptamer probe and FITC-labeled anti-CD30 antibody at room temperature for 20 min. As shown in Figures 5a–d, both the CD30 aptamer and anti-CD30 antibody independently bound to the CD30 proteins on the surface of Karpas 299 cells, suggesting that these two probes recognize different epitopes/regions of CD30 molecules, although the negative control Jurkat cells that do

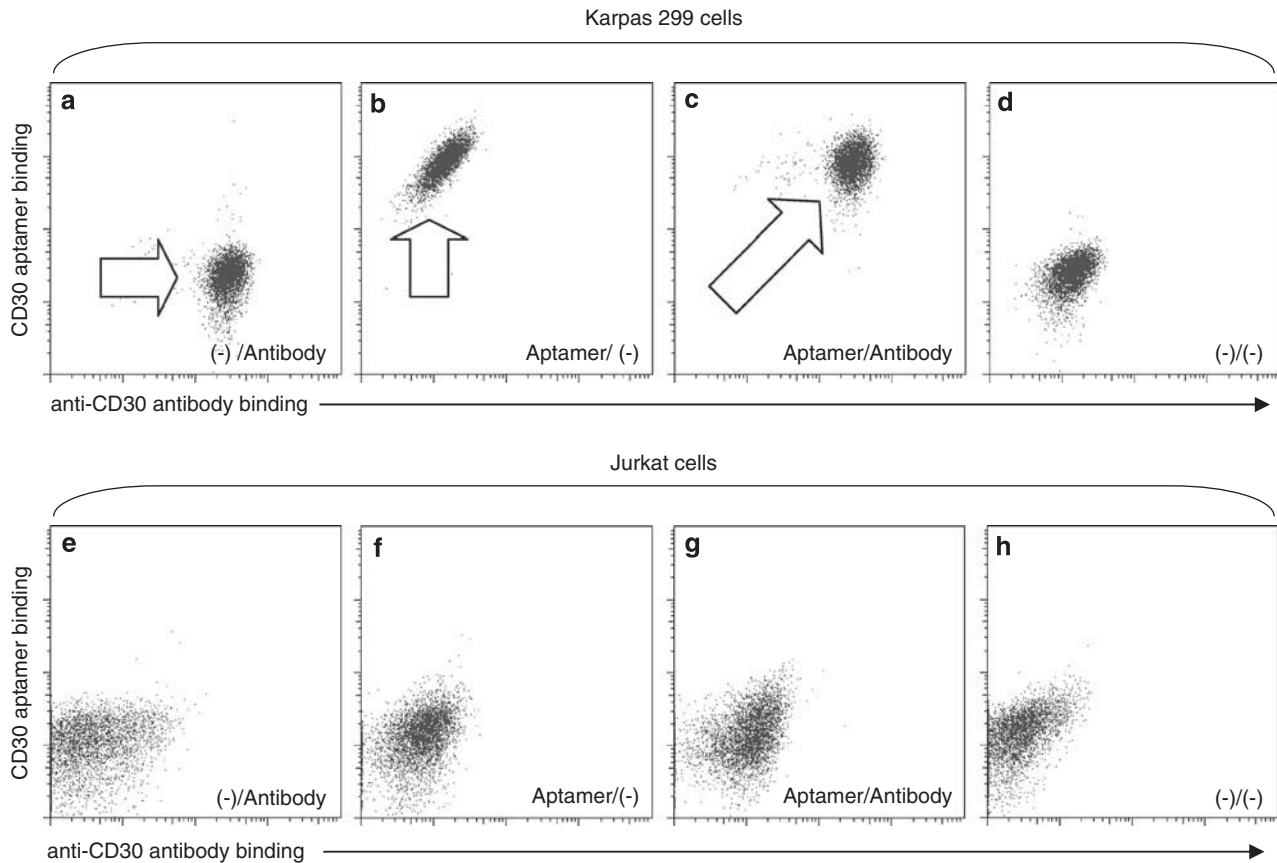


Figure 5 Double staining of cells by the CD30 aptamer and anti-CD30 antibody. Cultured Karpas 299 cells (a–d) and Jurkat cells (e–h) were incubated with the Cy5-labeled CD30 aptamer, FITC-labeled anti-CD30 antibody, or both simultaneously. Resultant cell staining was examined by flow cytometry analysis. Stained cell populations are indicated by arrows.

not express CD30 showed no label with either probes (Figures 5e–h).

Immunophenotyping of Cells by a Combination of the CD30 Aptamer and Antibodies

Advanced flow cytometry enables simultaneous detection of multiple biomarkers on individual cells. Specific type of cells could be identified and isolated by simultaneously using multiple specific probes that are labeled with different fluorochromes. Thus, it is interesting to determine whether the CD30 aptamer probe could be used in combination with other antibodies for multicolor cell staining. To answer this question, cells were incubated with a staining solution consisting of the Cy5-labeled aptamer probe specific for CD30 (a cell marker for ALCL and Hodgkin's lymphoma), and with a PE-labeled antibody specific for CD25 (a lymphocyte activation marker that is expressed by activated T and B cells as well as ALCL cells), AmCyan-labeled antibody specific for CD45, or FITC-labeled antibody specific for CD30. As shown in Figures 6a–f, Karpas 299 cells and SUDHL-1 cells were simultaneously stained by the CD30 aptamer probe and antibodies against CD25, CD30, and CD45. In the control group, Jurkat cells showed no staining by antibodies to CD25

or CD30 (both aptamer and antibody probes), but positive staining by the CD45 antibody as expected (Figures 6g–i).

Detection of ALCL Cells by the CD30 Aptamer Probe in a Mixed Cell Specimen

To validate whether the CD30 aptamer probe could be used in detecting ALCL cells from a mixed background of cells, a process more typical of the flow cytometry immunophenotyping for clinical lymphoma diagnosis, we used fresh bone marrow nucleated cells from healthy donors spiked with CD30-positive ALCL cells (Karpas 299). The samples were prepared and stained simultaneously with the Cy5-labeled CD30 aptamer, AmCyan-labeled anti-CD45 antibody, and FITC-labeled anti-CD30 antibody as described in 'Materials and Methods.' The populations of nucleated RBCs, blasts, lymphocytes, granulocytes, monocytes, and Karpas 299 cells were separated and gated according to their distribution on the side scatter (SSC) vs CD45 panels (Figure 7a). Cellular CD30 expression in each gated cell population was then evaluated by the CD30 aptamer or anti-CD30 antibody vs CD45 panels. As illustrated in Figures 7b and c, the CD30 aptamer and anti-CD30 antibody showed nearly identical staining patterns and numbers for CD30-

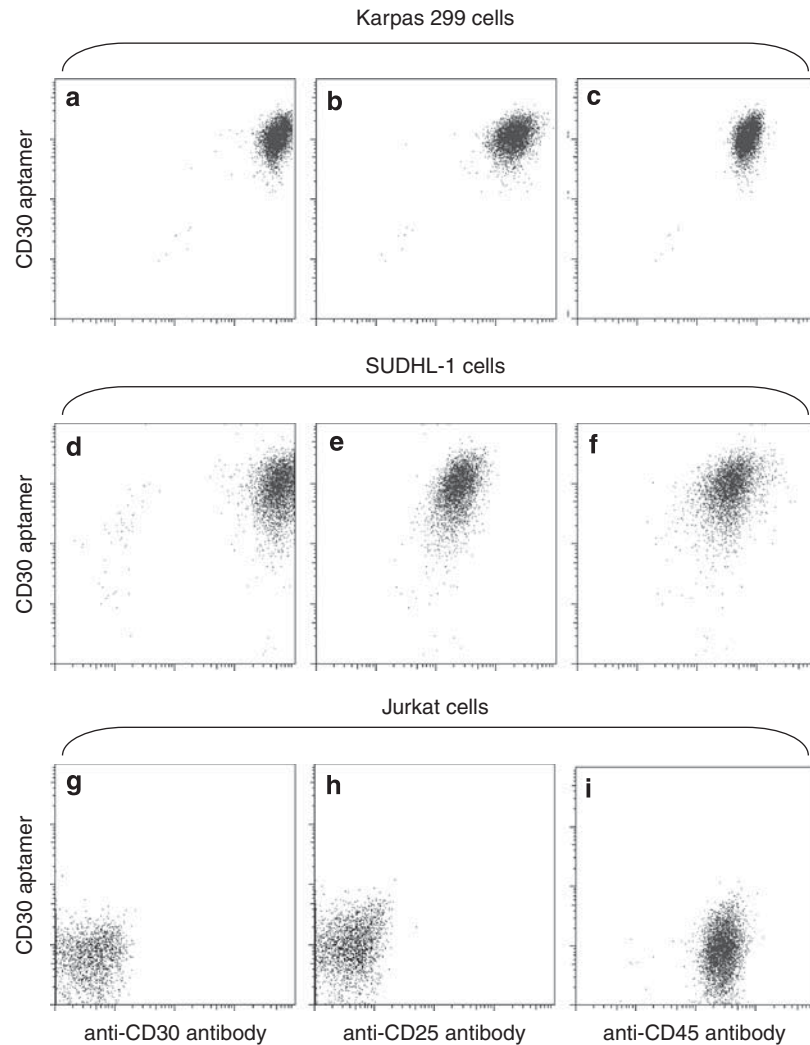


Figure 6 Multicolor staining of cells by combined exposure to the CD30 aptamer and antibodies. Cultured Karpas 299 cells (a–c), SUDHL-1 cells (d–f), and Jurkat cells (g–i) were incubated with mixed probes, including the Cy5-labeled CD30 aptamer, PE-labeled anti-CD25 antibody, FITC-labeled anti-CD30 antibody, and AmCyan-labeled anti-CD45 antibody. Resultant cell staining was examined by multicolor flow cytometry analysis.

positive Karpas 299 cells as well as CD30-negative background cells.

DISCUSSION

Currently, antibodies are generally the only available clinical probes for lymphoma diagnosis.²⁵ The study has shown that an RNA aptamer possesses identical specificity and sensitivity to those of anti-CD30 antibodies. Our results suggested that the CD30-specific aptamer could replace the anti-CD30 antibody in flow cytometry analysis. In addition, multicolor staining of cultured cells indicates that the CD30 aptamer probe can be used in combination with antibodies in multiparameter flow cytometry analysis for lymphoma diagnosis. It will be interesting to explore the use of CD30 aptamer probes for immunohistochemical staining of fixed tissue sections for the diagnoses of CD30-associated diseases, including ALCL and Hodgkin's Lymphomas. Before clinical application of the CD30 aptamer probe, a large-scale

validation study with samples from both normal donors and lymphoma patients should be performed.

In comparison with antibodies, oligonucleotide aptamers, such as those synthesized in this work, have unparalleled advantages for clinical use. As a small molecule, aptamers are not antigenic and can be synthesized at a significantly lower cost than antibodies.²⁶ In addition, aptamers can be easily modified and labeled with fluorescent, optical, or radioisotope reporters as needed for tracking purposes.^{4,6,7,23} These features increase the potential of aptamers for the *in vivo* application as diagnostic/therapeutic agents. Our findings indicate that the aptamer probe can be used in combination with antibodies for multicolor cell immunophenotyping analysis, and it may be a suitable replacement for the anti-CD30 antibody in disease diagnosis (Figure 7). In addition, studies using antibodies have revealed that the binding of the CD30 receptors results in the activation of the cellular signaling pathways that lead to the growth arrest and

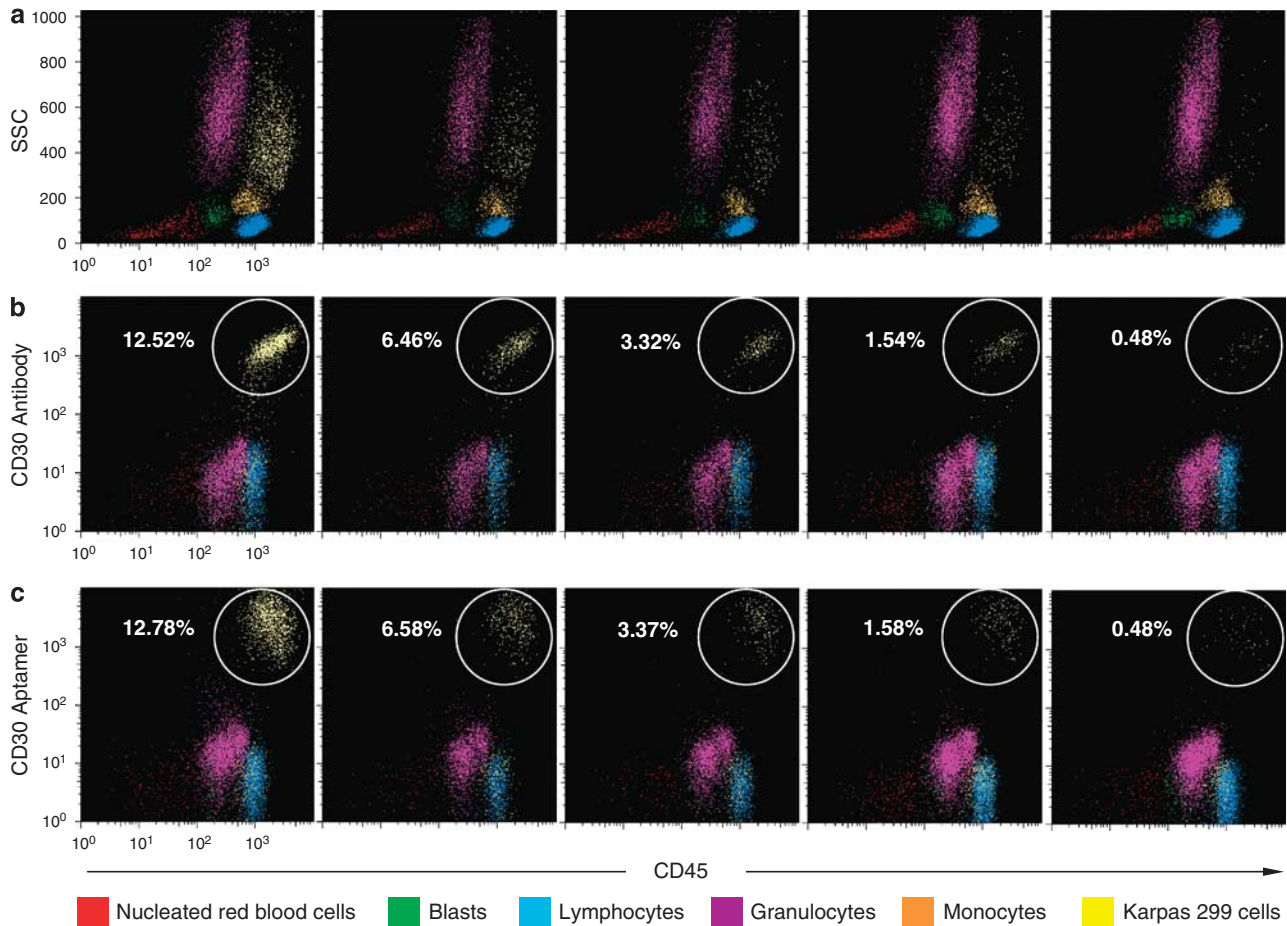


Figure 7 Detection of ALCL cells by the CD30 aptamer probe in a mixed cell specimen. **(a)** A mixture of fresh bone marrow nucleated cells and cultured Karpas 299 cells was stained simultaneously with the CD30 aptamer probe, anti-CD45 antibody, and anti-CD30 antibody. The populations of nucleated RBCs, blasts, lymphocytes, granulocytes, monocytes, and admixed Karpas 299 cells were separated and gated (painted in a different color as listed on the bottom) according to their distribution on the side scatter (SSC) vs CD45 panels. **(b)** CD30 expressions of individual cell populations gated in **(a)** were then examined by the anti-CD30 antibody vs CD45 panels and the numbers of detected CD30-positive Karpas 299 cells were indicated in each sample. **(c)** CD30 aptamer probe vs CD45 panels and the numbers of detected CD30-positive Karpas 299 cells.

death of ALCL cells *in vitro*.^{20,27–29} These findings suggest that the CD30 receptors may also be a therapeutic target for ALCL treatment. To understand whether the binding of aptamers to the CD30 receptors has any intracellular effect, we have exposed cultured ALCL cells (Karpas 299 and SUDHL-1) to the CD30 aptamer; however, the synthesized aptamer had no effect on cell growth as assessed by the MTT assay (unpublished result). It is notable that in contrast to antibodies that contain two binding sites to the CD30 receptors, the CD30 aptamer probes have only a single binding site that is not able to cross-link and activate the CD30 receptors on the cell surface.

Aptamers, especially the RNA-based aptamers, are susceptible to degradation by environmental nucleases, largely limiting their clinical value to date. However, chemical modification with 2'-O-methyl or 2-fluoro nucleic acids, phosphorothioate linkages, or a combination of these has been reported to significantly improve the nuclease resistance of the RNA-based aptamers.^{30–34} Considering the rapidly advancing synthesis technologies, we believe that aptamers

could become powerful diagnostic and therapeutic agents in the near future.

ACKNOWLEDGEMENTS

This work was supported by the National Institutes of Health (grant 5K22CA113493 to YZ), BCM-Methodist lymphoma SPORE (developing projects awarded to YZ and CHT), and TMHRI scholar and Clinical Translation Study Award (to YZ). Funding for open access charge: the NIH grant 5K22CA113493.

DISCLOSURE/CONFLICT OF INTEREST

The authors declare no conflict of interest.

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