

## Cell Penetrating or “Trojan” Peptides

During the last ten years it has been observed that a number of peptides and proteins are able to penetrate the cell membrane and enter the cell. Furthermore, it has been shown that even many cargo molecules that are covalently attached to these peptides will be translocated into the cell. Peptides that show the ability to translocate through the cell membrane are usually short peptides of less than 30 amino acids. Their only common feature appears to be that they are amphipathic and have a overall positive net charge. The exact mechanism of cell translocation is not known but appears to be receptor and energy independent, although in some cases their translocation can be partially mediated by endocytosis. The penetration into cells is usually rapid and of first-order, with half-times from 5 to 20 minutes [Zorko, M., and Langel, U., 2005].

The ability of a 60-amino acid polypeptide corresponding to the sequence of the *Drosophila* antennapedia gene homeobox to traverse across cell membranes of nerve cells and accumulate in the nuclei was first reported by Joiliet et al, in 1991. A shorter peptide coined penetratin is a 16 amino acids long peptide of cationic nature containing the sequence: **RQIKIWFQNRRMKWKK**. This sequence derives from helix 3 of the antennapedia complex [Thoren, P.E., et al., 2003; Fischer, P.M., et al., 2000] and is able to translocate through the plasma membrane to the cytosol and nucleus of living cells, both at 37 °C and 4 °C respectively.

Recently, methods have been developed for the delivery of exogenous proteins into living cells with the help of membrane-permeating carrier peptides derived from HIV-1 Tat (residues 48 to 60) and antennapedia (residues 43 to 58), penetratin [Derossi, D., et al., 1998; Dunican D.J., et al., 2001]. The basic nature of these peptides and the locations of aromatic groups within their sequence allows these peptides to penetrate the cell membrane. Investigating a range of basic peptides, Futaki reported that a peptide containing eight arginine residues can efficiently translocate across the cell membrane [Futaki, S. 2002]. Recently, a chimeric peptide derived from galparan and transportan has been used as an effective peptide vector for biodelivery of PNA molecules [Pooga, Marcus., et.al., 1998]. Translocation of the penetratin peptide occurs even when it is coupled to hydrophilic molecules (e.g. phosphopeptides, oligonucleotides, peptidic nucleic acids, drugs, etc.) [Prochiantz, A., 1996]. Taking the “*Illyad*” a tale from the Greek mythologies as an example these cell-penetrating peptides (CPP) were also termed ‘Trojan’ peptides. Most of them are water-soluble peptides with a low lytic activity that can be used as vectors for cellular internalization of hydrophilic biomolecules and drugs [Lindgren, M., et al., 2000, Stephens, D.J., & Pepperkok, R. 2001]. Despite its broad application field, the internalization mechanism of the penetratin peptide has not been totally unraveled yet. It appears that a receptor or a transporter protein is not needed, as retro-enantio and retro-inverso analogs of penetratin are also internalized [Derossi, D., et al., 1996]. Presumably, cellular internalization of the penetratin peptide occurs via a direct interaction with the cell membrane [Prochiantz, A., 1996].

Table 1 (below) contains a list of peptides that have been investigated for their ability to penetrate the cell. The potential to inhibit specific mRNA export pathways using cell-permeable peptides is shown in figure 1. Cell-penetrating peptide inhibitors are inhibiting their target proteins rapidly. In that sense they could be superior to other “functional knockout” approaches such as RNA interference (RNAi). RNAi prevents translation of a protein by destroying its mRNA; however, a long-lived protein will continue to be active long after its synthesis has stopped. The more rapidly a peptide affects RNA export, the more likely it is that the peptide directly (rather than indirectly) inhibits export of the target RNA. It is conceivable that designer inhibitory peptides based on the

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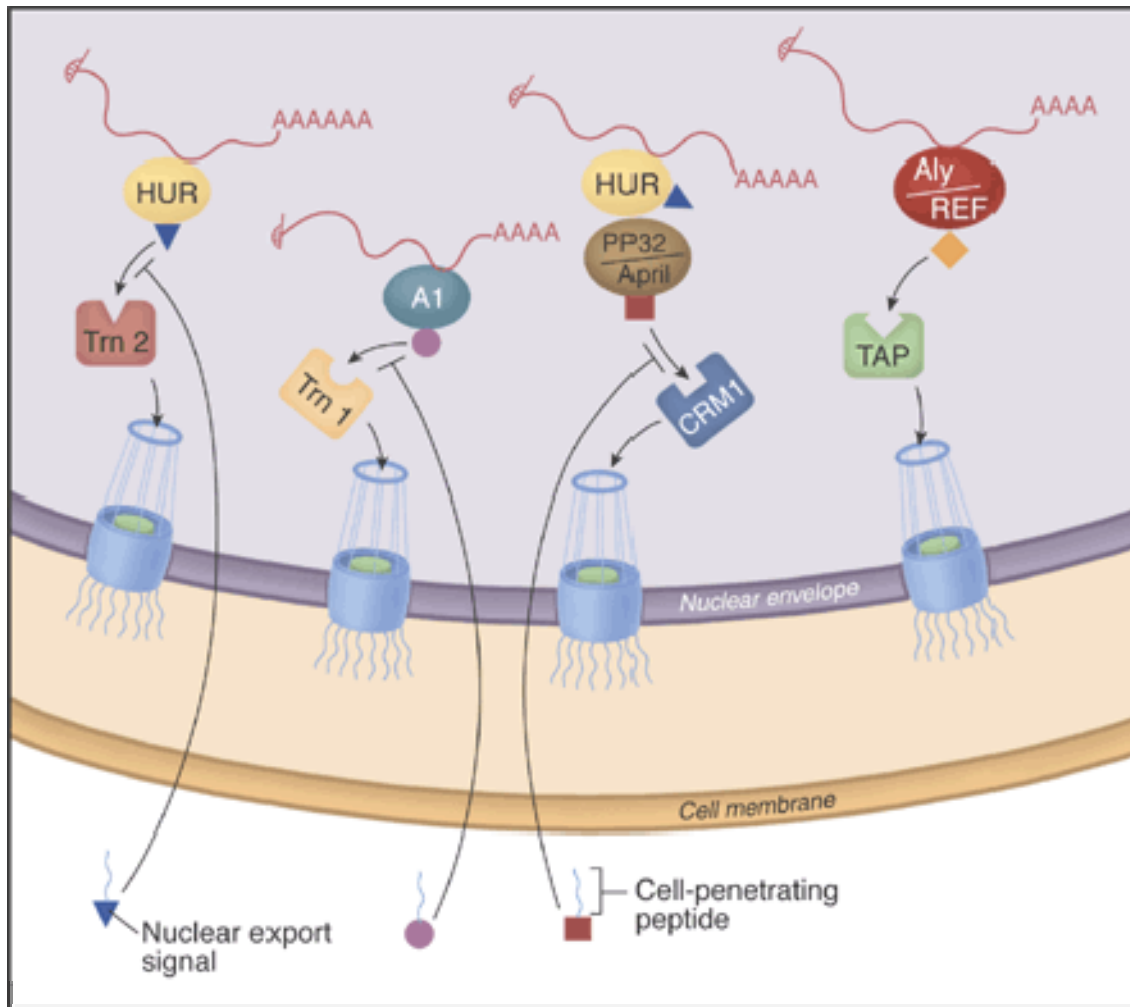
sequences of cell-penetrating peptides have the potential to allow for the generation of a mammalian cell tool kit that could parallel the temperature-sensitive mutant collection available to yeast geneticists [Moore and Rosbash, 2001].

**Table 1: Peptides that translocate into the cell** (Futaki, S. 2005)

Classes	Peptides	Sequences	Translocation efficiency
	<i>Tat and related peptides</i>		
<i>Basic</i>	HIV-1 Tat (48–60) Rn (n=7–11)	G <b>RKKRRQRRR</b> PPQ	+++
	D-Tat	G <b>RKKRRQRRR</b> PPQ	+++
	R9-Tat	G <b>RRRRRRRRR</b> PPQ	+++
	<i>Arginine-rich RNA binding peptides</i>		
	HIV-1 Rev-(34–50)	T <b>RQARRNRRRW</b> ERQR	+++
	R7W	Fluo- <b>RRRRRRR</b> W-NH <sub>2</sub>	+++
	TatP59W	Fluo-G <b>RKKRRQRRR</b> PWQ-NH <sub>2</sub>	+++
	FHV Coat-(35–49)	<b>RRRRNRTRNRRRVR</b>	
	BMV Gag-(7–25)	<b>KMTRAQRRAAAARN</b> WTAR	+++
	HTLV-II Rex-(4–16)	<b>TRRQTRRARNR</b>	+++
	CCMV Gag-(7–25)	<b>KLTRAQRRAAAARKNKR</b> NTR	++
	P22 N-(14–30)	<b>NAKTRRHERRRK</b> LAIER	++
<i>Basic/amphiphilic</i>	Antennapedia (43–58) ( <i>penetratin</i> )	<b>RQIKIWFQNRRMKWKK</b>	+++
	Fluoro-Penetratin	Fluo- <b>RQIKIWFQNRRMKWKK</b> -NH <sub>2</sub>	
	Pen2W2F	Fluo- <b>RQIKIFFQNRRMKF</b> KK-NH <sub>2</sub>	
	model amphipathic peptide	<b>KLALKLALKALKAAL</b> KL A-NH <sub>2</sub>	
	PenArg	Fluo- <b>RQIRIWFQNRRMR</b> WRR-NH <sub>2</sub>	
	PenLys	Fluo- <b>KQIKIWFQNKKMK</b> WKK-NH <sub>2</sub>	
	E N-(1–22)	MDAQT <b>RRRE</b> ERRA <b>EK</b> QAQW <b>K</b> AAN	+
	B 21 N-(12–29)	T <b>A</b> K <b>T</b> RY <b>K</b> ARRA <b>E</b> LIA <b>E</b> RR	+
	Yeast PRP6-(129–144)	T <b>RRNKR</b> NR <b>I</b> Q <b>E</b> QLNR <b>K</b>	+
	Hum U2AF-(142–153)	SQMT <b>RQARR</b> LYV	-
<i>Chimera</i>	<i>transportan</i> (galanin/mastoparan)	GWTLNSAGYLL <b>G</b> INL <b>K</b> ALAAL <b>A</b> KKIL	
	Pep-1 (hydrophobic/NLS)	<b>K</b> ETWW <b>E</b> TWW <b>T</b> EWSQP <b>KKKR</b> KV-cysteamine	
<i>Hydrophobic</i>	membrane translocating sequence peptide(MTS)	AAVALLPAVLLALLP	

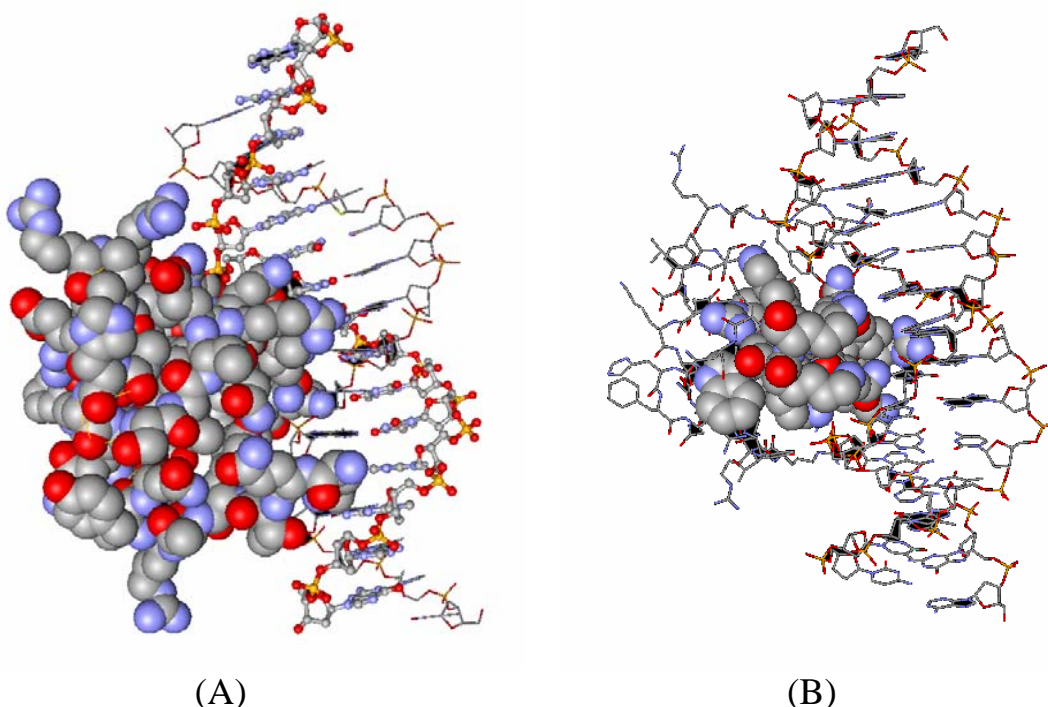
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Residues highlighted in dark bold blue indicate a positive charge where as the ones highlighted in bold red indicate negative charges.

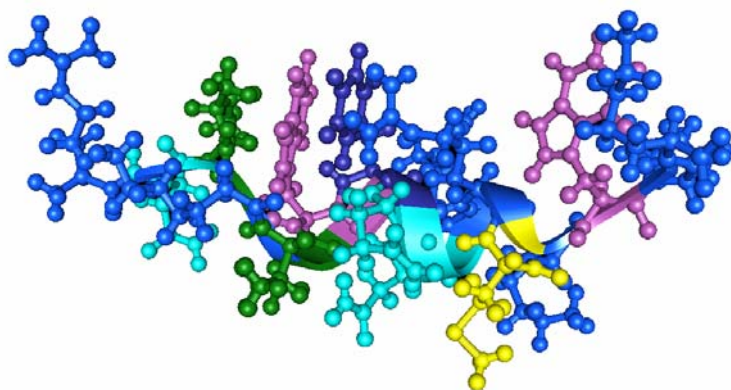


**Figure 1: The potential to inhibit specific mRNA export pathways with cell-permeable peptides.** The major route of mRNA export from the nucleus may depend on interactions between the mRNA adaptor protein Aly/REF and the export receptor heterodimer TAP:p15, which in turn interacts with the nuclear pore complex. New inhibitors that couple cell-penetrating peptides with specific nuclear export signals reveal that some mRNAs use other adaptors and receptors. For example, there are two nuclear export pathways for c-fos mRNA. One pathway involves the adaptor protein HuR and the export receptor Trn2, whereas the other involves HuR, its two ligands (pp32 and APRIL) and the export receptor CRM1. [Melissa J. Moore and Michael Rosbash in SCIENCE VOL 294 p. 1841, 2001].

## 3D structures



**Figure 2: Three-dimensional structure of the antennapedia homeodomain protein-DNA complex.** (Protein Data Bank accession code 9ant). A: The protein is shown as the space-fill model whereas the DNA is displayed in ball-and-stick (one DNA strand) and stick mode (second DNA strand). B: Only the peptide corresponding to penetratin peptide is now displayed in the space-filling mode all other atoms are in the stick mode. It is apparent that the peptide is in close contact with the DNA molecule apparently binding to the major groove of the helix.



RQIKIWFQNRRMKWKK

**Figure 3: Three-dimensional solution structure of penetratin.** (Protein Data Bank accession code 1kz0). Amino acid side chains of basic amino acid residues are highlighted in *blue*. The tryptophans are shown in purple. The primary structure of the peptides is also provided. All pictures were generated with the 3D mol viewer of Vector NT (version 6, Invitrogen, USA).

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## Cell Penetrating Peptides

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